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Monitoring of the Immune Response Activity in Iraqi Patients Infected with Cutaneous Leishmaniasis by IFN-γ and MIG Evaluation During Different Stages of Infection

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Abstract

Cutaneous leishmaniasis (CL) is one of the most prevalent cutaneous parasitic protozoan infections in Iraq; characterized by a chronic infection and granulomatous disease that invades the skin. Type 1 immune was predominates in CL patients with exacerbated production of pro-inflammatory cytokine, therefore this study aimed to evaluate serum level of interferon gamma (IFN- γ) and monokine induce by interferon gamma (MIG/CXCI9) as a useful markers of disease development in patients during different stage of infection (<1 month .. early , 1-6 month.. chronic and >6 months.. late). The result showed that there was an early effort to eliminate the parasite proliferation which illustrated by a high significant increase of both IFN- γ and MIG during the early stage of infection, but this response was down-regulated during the chronic stage of infection, which observed by low levels of both studied cytokine and monokine, nevertheless this down-regulation was transient where the levels to increase returns during the late stage of infection. The evaluation of IFN- γ and MIG considered as a biological markers of disease activity in each stage of infection.

Keywords: Cutaneous leishmaniasis, IFN-7, MIG, stage of infection.

متابعه فعاليه الاستجابه المناعيه في المرضى العراقيين المصابين باللشمانيا الجلديه بواسطه تقييم خلال مراحل المرض المختلفة MIG وγ-IFN

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الخلاصة

مرض اللثمانيا الجلدية هو احد اكثر الاصابات الجلدية الطفيلية الابتدائية في العراق. يتمثل بالاصابة المزمنة والورم الحبيبي الذي يغزو الجلد. النوع الاول من الاستجابه المناعيه في مرض اللثمانيا الجلديه تكون هي السائده مع تفاقم في انتاج المدورات الخلويه البادئه بالالتهاب ولهذا هدفت الدراسه الحاليه الى تقييم مستوى γ -NF و MIG في المصل كدلائل مفيده لتطور المرض في المرضى خلال مراحل مختلفه من الاصابه (اقل من شهر ..مبكره ، 1-6 مزمنه ومتاخره اكثر من سته اشهر). اشارت النتائج الى وجود جهود مبكره للحد من تكاثر الطفيليات والتي تمثلت بارتفاع معنوي في كلا γ -NF و MIG خلال المرحله المبكره من الاصابه، لكن هذه الاستجابة تتاقصت خلال مرحله المرض المزمنه والتي توضحت بنقصان مستوى كلا العاملين المدروسين، ومع ذلك هذا النتاقص كان عابر وذلك لعوده ازدياد كلا العاملين الى الارتفاع خلال المرحله المتاخره من المرض. قياس FN–γ و MIG يعتبر كمؤشرات حيويه لفعالية و شده المرض في كل مر حله من مر احل الاصابة.

Introduction

Leishmaniasis is a severe parasitic disease, viewed via the World Health Organization (WHO) to be one of the most necessary ignored diseases globally. Approximately 12 million people are infected with greater than twenty distinct pathogenic *Leishmania* species worldwide (i.e. tropical and subtropical areas of Asia, the Middle East, sub-Saharan Africa, and South America) [1].

The disease is caused by protozoa belonging to the genus *Leishmania* and transmitted through the chunk of a 2–3 mm lengthy insect vector, the *Phlebotomus* and *Lutzomyia* sand fly [2]. The medical photo of leishmaniasis is heterogeneous with a large spectrum of human diseases [3]. *Leishmania* parasite can give rise to different form depending on the infecting species such localized cutaneous leishmaniasis (CL), CL with mucosal involvement (MCL), visceral leishmaniasis (VL) [4].

According to [5] CL is the most common form of the disease, with many different *Leishmania* species responsible for signs and symptoms from both the Old World (*L. major, L. tropica, L. aethiopica*) and New World (*L. mexicana, L. amazonesis, L. braziliensis, L. panamensis, L. guanensis*).

The immune response to CL is characterized by a mixed T-helper 1 and T-helper 2 immune response [6]. The Th1 immune response is necessary to manage parasite growth and to stop *Leishmania* proliferation and dissemination, [7]. Interleukin-2 (IL-2) and interferon-gamma (IFN-gamma) are usually produce in large amount by Th1 [8]. On the other hand, the Th2 immune response is responsible for development and chronicity of lesions in *leishmania* infection. Th2 cell characterized by production of IL4 and IL10 cytokine [9].

Interferon gamma (IFN-Y) is a major Th1 derived cytokine that plays crucial role in controlling *Leishmania* infection [6]. It involves in induction of cidal activity against *Leishmania* parasite and induces the production of oxygen species [10].

One of the important immunological mediators in prime inflammation is monokine induced by IFN-Y (MIG/CXCL9) which is the predominant function of it is the recruitment of primed T-lymphocyte to sites of inflammation [11]. This monokine is a member of the CXC subfamily of chemokine. The manufacturing of MIG is specifically activated via IFN-Y and can also play necessary roles in improving the recruitment and activation of Th1 cells [12], and stimulate T-lymphocyte proliferation and enhance T-lymphocyte effector cytokine production [11].

Materials and methods

Subject selection

This study included two groups of individual:

- First group included sixty-six patients infected with cutaneous leshmaniasis collected from AL-Karama Hospital in Baghdad (42 male and 26 female), their ages ranged from 1 year to 55 years during the period November 2018 to February 2019.

- The second group includes forty healthy individuals, as a control group (22 male and 18 female), their ages ranged from 5 to 68 years.

All study participants signed the informed consent form agreeing to participate in the study. Questionnaire sheet was taken for each individual includes: Name, sex, age, duration of infection, the number of lesions, and treatment.

Blood sample collection

Five ml of venous blood was collected from each patient with a disposable syringe. The collected samples were placed in a vacuum tubes and left to stand at room temperature to clot formation after that the tubes were centrifuged at 3000 rmp, then the sera were collected in Eppendorf tubes and stored at -20 °C until used.

Immunological assays

Determination of IFN- γ

Measurements of IFN- γ were performed according to the direction of the manufacturer of Abcam ELISA (ab100537) kit.

Determination of MIG

Measurements of MIG were performed according to the direction of the manufacturer of MyBio Source ELISA kit.

Statistical analysis

The Statistical Analysis System- SAS [13] program was used to detect the effect of difference factors in study parameters. Least significant difference –LSD test (Analysis of Variation-ANOVA) and T-test was used to significant compare between means.

Results and discussion

Cutaneous leishmaniasis

Sex distribution

In this study the total numbers of cutaneous leishmaniasis patients were 66 individuals, 40/66 (60.6 %) males and 26/66 (39.39 %) females.

Age distribution

The ages of patients studied were ranged between 1 years to 55 years, the majority of patients were at the age of <15 years 28 (42.42%).

Body site of infection

Different body parts of patients were found to be infected with CL which includes face, arms and legs, but the highest percentage was observed in arm 29/66 (43.93%) following by legs 23/66 (34.84%) and face 14/66 (21.21%), Table-1.

Duration of infection

Duration of infection with CL in patients was ranged from <1 months to >6 months, Table-1. **Number of lesions**

The number of lesions ranged between 1-18 in different body part,25/66 (37.87%) of CL patients had one ulcerated lesion, 41/66 (62.12%) with multiple ulcerated lesions, Table-1, Figure-1.

Table 1- The percentage of CL distribution according site of infection, duration of infection and number of lesions.

		No. of case (%)	Total no. (%)
	Arms no. (%)	29(43.93)	
Site of infection	Legs no. (%)	23(34.84)	66(100)
	Face no. (%)	14(21.21)	00(100)
Duration of	Early infection	35(53.03)	
infection	Chronic infection	27(40.90)	66(100)
	Late infection	4 (6.06)	00(100)
Number of lesions —	One lesion	25(37.87)	
inumber of lesions	Multiple lesions	41(62.12)	66(100)



Figure 1-Patient with multiple lesions on face

Serological findings by ELISA Gamma-interferon (IFN-γ)

The results of this study obtained that patients with CL had significant increased (p<0.01) of IFN- γ mean level (40.94 ± 1.17 pg/ml) in comparison to control group (3.04 ± 0.53 pg/ml), Table-2. **Table 2-**Mean level of IFN- γ in CL patients in comparison to control group

Group	Mean ± SE (pg/ml)	T-test	P-value
Patients	40.94 ± 1.17	3.377 **	0.0001
Control	3.04 ± 0.53	5.577	0.0001
** (P<0.01).			

This result was agreed with previous results by [14] who showed significant increase of IFN- γ level in CL patients in comparison to control group while this result was disagreed with Al-Aubaidi [15] that obtained decreased level of IFN- γ in CL patients in comparison to control group. These differences may relate to different kits or different type of sample or the strain of the parasite; but in fact the current result was tending to be more truth, especially when there is an early infection there is an acute inflammation so there is a high inflammatory cytokine level in comparison to control group. At the site of parasite entrance the cutaneous leishmanial lesions take place and begin as a small

papule that develops into a nodule that ulcerates in the center [16].

This increase in IFN- γ level in the current study is required to eliminate the intacellular parasite. Cell mediated immune responses plays an important role in control intra cellular infection and resolve disease of leishmaniasis [8]. Furthermore, resistance to infection or recovery is related with a Th-1 response that is based upon expression of IL-12, IFN- γ , and TNF- α with the production of nitric oxide (NO) [17]. Interferon gamma is a Th1 cytokine that plays an important role in limiting the growth of *leishmania* parasite and controlling the progression of disease by enhancing macrophages activity [18].

The current results also showed that patients with early CL (< 1 months) had the highest significant increased (p<0.05) mean level (44.05 \pm 2.19 pg/ml) of IFN- γ than other patients with different duration of infection. However, late CL (> 6 months) (40.40 \pm 2.68 pg/ml) mean level showed higher significant (P<0.05) in comparison to chronic stage (1-6 months) (37.81 \pm 1.12 pg/ml), Table-3.

Table 3- Mean concentration of IFN- γ in CL patients according to duration of infection, number of lesion and treatment.

	Group	Mean ± SE (pg/ml)	LSD value
	Early infection	44.05 ± 2.19	
Duration of – infection –	Chronic infection	37.81 ± 1.12	
	Late infection	40.40 ± 2.68	6.016 *
Number of	One lesion	45.25 ± 2.37	
lesions	Multiple lesions	38.79 ± 1.10	4.631 **
Treatment –	With treatment	44.60 ± 1.69	
	Without treatment	36.69 ± 0.85	4.084**
* (P<0.05), ** (P<0.01), NS: Non-Significant.			

This results was in agreement with Bacellar *et al.* [19] who showed that IFN- γ and TNF- α were present at higher concentrations during the primary *Leishmania* infections when compared to controls. These cytokines activate macrophage that plays a crucial role in parasite killing.

Also, Ajdary *et al.* [20] said that a high level of IFN- γ production was observed to be associated with spontaneous lesion cure. Furthermore, chronic lesions and therapeutic failure had been associated with low level of IFN- γ production.

Another studies confirm this fact, Castellano *et al.* [6] study that the antibody plasma levels and cytokine production in peripheral blood mononuclear cells (PBMC) culture supernatants in patients with active or healed lesions and revealed that higher IFN- γ levels was observed in patients with active lesions and its persistence at elevated levels after lesion healing. Also Handjani *et al.* [10] demonstrated that the clinical healing of lesion of CL has been associated with Th-1 immune response

characterized with high level of IFN- γ production and decreased of Th-2 immune response and its related cytokine such IL-10.

On the other hand, the current results was disagreed with Ajdary *et al.* [21] showed that the level of IFN- γ was similar between early and persisted CL patients but it was higher in comparison to control group. They concluded that non-healing patient of CL shows a mixed of Th1/Th2 responses whereas patients with acute lesion presents a Th1 response.

During the primary stage of infection the innate immune system of the host must quickly detect and respond to *leishmania* parasite infections, and this is performed by innate immune receptors. In the absence of recognition by innate immune receptors, the host will quickly become overwhelmed by parasite pathogens. On the other hand, if activation of innate immune receptors is too much, high levels of pro-inflammatory mediators such as TNF- α and IFN- γ could be detrimental to the host [22].

The down-regulation of the immune response in CL patients is a transitory phenomenon for the reason that virtually all patients with more than 6 months of history of cutaneous lesion present strong IFN- γ production. Also, this down-regulation of IFN- γ production in CL can also account for parasite multiplication.

Also, the mean level of IFN- γ cytokine observed significantly (p<0.01) high increase mean (45.25 ± 2.37 pg/ml) in a single lesion patients in comparison to patients with multiple lesions, mean (38.79 ± 1.10 pg/ml), Table-3.

Galgamuwa *et al.* [23] found that the expression of IFN- γ cytokine was higher in single lesions compared to people having multiple lesions of CL. Furthermore, these findings confirmed that levels of IFN- γ was higher in serum patients with primary infection as compared with sera obtained from patients with secondary *Leishmania* infection [17]. These results were in line with the present result.

The explanation of this increase may relat to the initial infection always characterized by high proinflamatory cytokines as IFN- γ , but when there is a secondary and tertiary etc. infection (ulcers) the immune response modulated towards Th-2 and their cytokines such as (IL-10, IL-4, TGF- β etc.) so that level of IFN- γ tend to decrease and the parasite were increased in their intensity in multiple lesion. The results also showed that IFN- γ level was significantly (P<0.01) higher in treated patients mean (44.60 ± 1.69 pg/ml) in comparison to patients without treatment mean (36.69 ± 0.85 pg/ml), Table-3.

This results was consistent with preveous result that revealed a significant increase in IFN- γ level during treatments in comparison to its level before treatment and this clarified that a successful treatment was activated T-cell proliferation in response to *leishmania* antigen [15].

Also, another finding confirms this fact, increased IFN- γ production at the end of the treatment in human is due to the role of this cytokine in healing process and involvement of CD4+ T-cell in this process [24].

Monokine induces by interferon gamma MIG

The results of this study showed that patients with cutaneous leishmaniasis had higher significant (P<0.001) levels of total MIG mean (257.08 \pm 8.54pg/ml) in comparison to the control group mean (220.83 \pm 5.16 pg/ml), Table-4.

Group	Mean ± SE (pg/ml)	T-test	P-value
Patients	257.08 ± 8.54		
Control	220.83 ± 5.16	26.155 **	0.0071
** (P<0.001).			

Table 4-Mean level of MIG in CL patients in comparison to control group

The current results was in agreements with previous results by Giudice *et al*, [25] who study peripheral blood mononuclear cell-derived macrophages which exposed to *L.braziliensis in vitro* and evaluated for susceptibility to the infection, the ability to kill *Leishmania* parasite and chemokine/cytokine production and their result found that macrophages from CL patients produced more chemokines and TNF- α than those from the healthy control group and the levels of MIG were significantly higher in CL.

Lesions of CL is characterized by an exacerbated cell-mediated immunity that forms a granuloma which contain many activated T-cells and produces a variety of cytokines [26]. The regulation of cellular migration from lymph organ to peripheral tissues plays a crucial role in the immune response. Chemokine and chemokine receptors are critical to the molecular mechanisms driving in this

procedure. A combination of adhesion molecules and chemokine receptors mediate the recruitment of naive and memory T cells to peripheral tissue[27].

Cytokines are directly involved in chemokine manufacturing and can also precede the expression of chemokines. Th1-related chemokines such as induced protein-10 (IP-10) and monocyte chemoattractant protein(MCP) is induction by IL-12, and interferon (IFN)- γ selectively induces IP-10 and MIG [28].

In *Leishmania* infection the role of chemokines has been exhaustively studied and these findings indicate that chemokines and chemokine receptors play an important role in detictive the outcome of the disease [29].

Additionally, MIG is secreted in high levels in auto-immune diseases such multiple sclerosis [30] and chronic inflammatory like human T cell lymphotropic virus infection. In both cases, MIG is associated with inflammation and the disease pathology [31].

The results as well showed that patients with early CL (<1 month) mean (279.55 \pm 13.28) had highest mean levels of MIG than other patients with different duration of infection, however late CL (>6 months) mean (241.75 \pm 42.87pg/ml) showed higher level in comparison to (1-6 months) of infection but this result was non-significant, Table-5.

Moreover the results obtained that the level of MIG was lower in patients who had 1 lesion on their body mean (256.17 ± 14.76 pg/ml) in comparison to patients who had multiple lesion mean (257.63 ± 10.54 pg/ml), but this result also was non-significant, Table-5.

Table 5-Mean concentration of MIG in patients of CL according to duration of infection, number of lesion and treatment

	Group	Mean ± SE (pg/ml)	LSD value
	Early infection	279.55 ± 13.28	
Duration of infection	Chronic infection	230.22 ± 7.84	60.575 NS
meetion	Late infection	241.75 ± 42.87	00.375 115
Number of	One lesion	256.17 ± 14.76	
lesions	Multiple lesions	257.63 ± 10.54	35.462 NS
	With treatment	281.27 ± 15.56	
Treatment	Without treatment	238.12 ± 8.04	32.943 **
** (P<0.01), NS: Non-Significant			

The present data was agree with Campanelli *et al.* [32] who detected that in the early stages of CL (<30 days of infection) there was a higher expression of chemokine receptore (CXCR3) ckemokines than in late stages of disease (>60 days of infection).

Moreover the results obtained that the mean level of MIG showed no significant difference (P>0.05)in patients who had one lesion on their body ($256.17 \pm 14.76 \text{ pg/ml}$) in comparison to patients who had multiple lesion ($257.63 \pm 10.54 \text{ pg/ml}$), Table-5.

The results also displayed that MIG mean level was significantly (P<0.01) decreased in patients without treatment in comparison to treated patients, Table-5.

Here in the present results the data showed an interestingly raised production of MIG during treatment in comparision to not treated one, this result was in agreement with previous result by Machado *et al.* [33] explained that MIG and IP-10 ckemokines were produced by the stimulation of *leishmania* antigen by clustered monocytic cells in the lesion.

Conclusion

From the current study the data investigated that there was an early effort to eliminate the parasite development which characterized by the high significant increase of IFN- γ level that play a role in recruitments of T-cell to the site of infection for cell-mediated immunity, this thought illustrated by the increased level of MIG. But these efforts were down-regulated over time during the period (1-6 months) which may result from the immune manipulation of the parasite within the host as one of the strategies followed for immune evasion.

The down-regulation of the Th-1 cytokine in the present study seems transient because CL patients with late stage of infection presented a high significant level of IFN- γ , MIG. This increased in Th-1 cytokine may lead to clinical healing of CL lesion.

Recommendation

Evaluation of the expression of the chemokines (CXCL10, CCL8, CCL9 and the chemokine receptors (CR3, CXCR3, CCR5) in the lesions of CL patients with different clinical forms by using immunohistochemistry. Studying the Immunohistochemical of skin biopsies to elucidate the cytokine release (IL-12, IL-4, and TNF- α) capabilities of immune cells and for immune-phenotypic patterns (CD4, CD8, CD14, CD19 and CD56).

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